



Combined zinc and nitrogen applications at panicle initiation for zinc biofortification in rice

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Abstract

Background and purpose: Increasing zinc (Zn) concentration in rice grains can help improve Zn nutrition of people. The combinations of Zn and nitrogen (N) applications at panicle initiation were investigated for Zn biofortification in rice.

Materials and methods: Rice (cv. Super Basmati) seedling were grown in pots having a calcareous soil. All combinations of four Zn (control, Soil 6 mg Zn kg⁻¹, foliar 2 × 0.2% Zn and soil + foliar Zn) and three N (control, soil 20 mg N kg⁻¹ and foliar 0.5% N) levels were imposed at panicle initiation. At maturity, grains analysed for Zn and proteins.

Results: Grain protein concentration was significantly increased with foliar Zn treatments, and with soil and foliar N. Maximum grain Zn concentration (30 mg kg⁻¹) was achieved with application of soil Zn + foliar Zn + foliar N. At each N level, Zn application by either method significantly increased grain Zn concentration over control. This increase in grain Zn concentration at N levels was 36 to 54% with soil Zn + foliar Zn, 27 to 45% with foliar Zn and 9 to 15% with soil Zn over its control level.

Conclusions: Grain Zn concentration was significantly increased with soil N when combined with soil Zn, and with foliar N when combined with foliar Zn treatments. Conclusively, foliar N combined with soil + foliar Zn is the best combination of late Zn and N application for agronomic Zn biofortification in rice.

INTRODUCTION

Lacking the affordability to a diversified food, people in many countries primarily rely on cereal grains to sustain their lives. Over time, intensive agriculture has depleted nutrients from soils (1) resulting in a poor nutritional quality of cereal grains produced from these soils. Moreover, the green revolution in the 1960s led to the development of high yielding cereal cultivars that have lower mineral density than old cultivars (2, 3). As a consequence, people are encountering undernourishment of iron (Fe), zinc (Zn) and several other nutrients.

Zinc deficiency causes human health problems such as poor physical growth, weak immune system, low learning ability and deoxyribonucleic acid damage. Zinc biofortification of staple crops, both genetic and agronomic, is a promising approach and it has gained due perception by Consultative Group on International Agricultural Research (4). Wheat, rice, and maize are important cereal crops that are the targets of Zn biofortification as being consumed mainly in countries having Zn deficiency in human populations. In rice grains, efforts are being made to

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increase the current baseline Zn level of 16 mg kg⁻¹ to an acceptable level of 28 mg kg⁻¹.

Alkaline calcareous soils and flooded field conditions in the rice production systems in Asia decrease the availability of soil Zn to growing rice plants (5). Not only the yield but also the quality of the produced grains is low under such production systems. Other factors contributing to low Zn availability from soil are low organic matter, high phosphate application and salt stress (6). Agronomic biofortification by Zn application may simultaneously increase grain yield and grain Zn concentration in both standard and biofortified cultivars of cereals (7). Zinc application by different methods significantly improved grain Zn concentration in rice (8–10). Grain Zn concentration is enhanced more with foliar application of Zn during flowering and grain development stages than other methods of Zn application to the rice crop.

Zinc in grains is localized with proteins (11). Application of N, both as basal and late dose, is known to increase root uptake, root-to-shoot translocation and remobilization of Zn (12). For Zn biofortification in rice, combined applications of Zn and N to rice are more important than their sole applications (13). However, it is still unknown if soil or foliar application of N at panicle initiation will be a better combination with different application methods of Zn to rice. The specific objectives of the experiment were to investigate: (i) the effect of late Zn and N applications on grain protein concentration and (ii) the best combination of Zn and N applications for Zn biofortification in rice.

MATERIAL AND METHODS

A pot experiment was conducted in a glasshouse at Department of Soil Science, Bahauddin Zakariya University, Multan (Pakistan). Soil for the study was collected from the surface layer (0–30 cm soil depth) of a field of at Agricultural Research Farm of the university. The soil was air-dried, crushed, and passed through a 2 mm sieve. A representative subsample of the soil was analysed for basic characteristics following standard methods (14, 15). The clay loam (determined by Hydrometer method) had sand, silt, and clay contents of 26, 40 and 34%, respectively. The soil was alkaline calcareous in nature having pH 7.8 in saturated soil paste and 4% w/w acid neutralizable free CaCO₃. The electrical conductivity of soil saturated paste extract (EC_e), organic matter content and DTPA-extractable Zn of soil were 2.4 dS m⁻¹, 0.5% w/w, 0.7 mg kg⁻¹, respectively.

Each of 36 polyethylene lined plastic pot was filled in with 10 kg of the soil. Rice (cv. Super Basmati) seedlings (25 d old) were purchased from the local market. Three pairs of healthy seedlings of uniform height were transplanted in each pot in the last week of July 2016. A basal dose of nitrogen (25 mg kg⁻¹ soil), phosphorus (25 mg kg⁻¹ soil) and potassium (31 mg kg⁻¹ soil) was applied as urea [(NH₂)₂CO] and potassium di-hydrogen phosphate

(KH₂PO₄), respectively. After transplantation, water level in the pots was kept about 3 cm above the soil for the first 14 days, and then progressively increased to 5 cm height. For this purpose, tube-well water of good irrigation quality (pH 7.8, EC 0.34 dS m⁻¹, and undetectable Zn concentration) was used. All pots were randomized every 6th day to avoid the differential effects of microclimate in the glasshouse. Second and third splits of N, each of 25 mg kg⁻¹ soil, were applied after 3 and 6 weeks of transplantation, respectively.

Experiment had twelve treatments in total, i.e. all possible combinations of four Zn levels (control, soil application at 6 mg Zn kg⁻¹, 2 foliar sprays of 0.2% w/v Zn solution and soil + foliar application) and three N levels (control, soil application at 20 mg N kg⁻¹ and foliar spray of 0.5% w/v N) were applied at panicle emergence stage of rice. These fertiliser treatments were of urea [(NH₂)₂CO] and hydrated zinc sulphate (ZnSO₄·7H₂O) and were applied additionally to the basal applications of macronutrient fertilisers. The treatments were arranged in completely randomized factorial design with three replications.

Irrigation was stopped a week before the harvest of the crop at maturity. Straws and panicles were cut and collected in separate paper bags. The plant samples were kept in an oven at 65 °C till constant weight and this was followed by the recording of straw and grains dry weights. Unhusked rice grains were dry ashed in a muffle furnace at 550°C followed by dissolution in 5 N hydrochloric acid (HCl) and dilution with distilled water (16). Zinc in the digests was analysed on an atomic absorption spectrometer (Thermo Scientific 3000 Series, Waltham, MA, USA). For determination of N, separate samples of unhulled grains were digested on a hot plate with sulfuric acid (H₂SO₄) and hydrogen peroxide (16). In wet-digested samples, a colour was developed with Nessler's reagent and absorbance was measured on a spectrophotometer (UV-1602, BMS, Quebec, Canada) 425 nm (17). A factor of 5.7 was used to convert total N measurements to concentration of raw protein in grains.

Statistical significance ($P \leq 0.05$) of main and interactive effects of treatments on the recorded parameters was tested by two-way analyses of variance (ANOVA) test with interactions. The significant difference among measured was determined by Tukey's HSD test. All statistical analyses were carried out on SAS University Edition (SAS/STAT®, SAS Institute Inc., NC, USA).

RESULTS

Rice yield

Main effects of Zn and N significantly ($P \leq 0.05$) influenced grain and straw yield of rice (Table 1). As compared to Zn-control, grain and straw yields were increased by Zn application to the soil (Table 2). With soil Zn alone and Soil Zn + foliar Zn, grain yield increased respective-

ly by 7 and 9% while that of straw yield by about 7%. Foliar-applied Zn did not affect straw or grain yield.

As compared to control level of N, grain and straw yields were increased by about 5% each with foliar-applied N and by 11 and 9%, respectively with soil-applied N (Table 2). In contrast to straw yield, grain yield was more with soil application of N than its foliar application.

Grain protein

Main effects of both Zn and N significantly ($P \leq 0.05$) influenced concentration and contents of raw protein in grains (Table 1). However, variance in concentration and contents of grain protein was contributed more by N than Zn application.

Soil application of Zn did not affect grain protein concentration while foliar application alone or in combination with soil Zn increased grain protein concentration by 4%

Table 1. Outcome (*F* values) of two-way analysis of variance (ANOVA) test

Parameter	Source of variation		
	Zn	N	Zn \times N
Grain yield	3*	11*	1
Straw yield	3*	9*	1
Grain protein concentration	4*	53*	1
Grain protein contents	5**	37*	1
Grain Zn concentration	161*	19*	6*
Grain Zn contents	65*	22*	2

Asterisk (*) denotes significant effect at $P \leq 0.05$.

Table 2. Yield response of rice grown in pots and fertilised, at panicle emergence stage, with all combinations of four zinc (Zn) levels and three nitrogen (N) levels

Treatment levels	Grain yield (g pot ⁻¹)	Straw yield (g pot ⁻¹)
Main effect of Zn levels		
Control Zn	5.5 \pm 0.4 B	16.1 \pm 1.2 B
Soil Zn (6 mg Zn kg ⁻¹)	5.9 \pm 0.2 A	17.0 \pm 0.8 A
Foliar Zn (0.2% w/v Zn)	5.7 \pm 0.4 AB	16.5 \pm 1.2 AB
Soil Zn (6 mg Zn kg ⁻¹) + Foliar Zn (0.2% w/v Zn)	6.0 \pm 0.5 A	17.1 \pm 0.9 A
Main effect of N levels		
Control N	5.5 \pm 0.3 C	15.9 \pm 0.9 B
Soil N (20 mg kg ⁻¹ soil)	6.1 \pm 0.3 A	17.4 \pm 0.7 A
Foliar N (0.5% w/v N)	5.8 \pm 0.4 B	16.7 \pm 1.2 A

Means \pm standard deviations; Separately for each main effect, different letters indicate significant ($P \leq 0.05$) differences based on Tukey's HSD.

Table 3. Concentration and contents of raw proteins in grains of rice grown in pots and fertilised, at panicle emergence stage, with all combinations of four zinc (Zn) levels and three nitrogen (N) levels

Treatment levels	Grain protein concentration (g kg ⁻¹)	Grain protein content (mg pot ⁻¹)
Main effect of Zn levels		
Control Zn	76 \pm 5 B	423 \pm 53 B
Soil Zn (6 mg Zn kg ⁻¹)	77 \pm 5 AB	457 \pm 43 A
Foliar Zn (0.2% Zn w/v)	79 \pm 4 A	455 \pm 43 A
Soil Zn (6 mg Zn kg ⁻¹) + Foliar Zn (0.2% Zn w/v)	79 \pm 4 A	469 \pm 54 A
Main effect of N levels		
Control N	73 \pm 3 B	399 \pm 28 C
Soil N (20 mg kg ⁻¹ soil)	80 \pm 2 A	489 \pm 29 A
Foliar N (0.5% N w/v)	80 \pm 2 A	464 \pm 38 B

Means \pm standard deviations; Separately for each main effect, different letters indicate significant ($P \leq 0.05$) differences based on Tukey's HSD.

each over control level of Zn (Table 3). Zinc application, by any method, increased grain protein contents over control level of Zn; the increase ranged from 8 to 11%. The differences in both concentration and contents of grain protein were non-significant ($P \leq 0.05$) at three applied levels of Zn.

Nitrogen application by either method significantly ($P \leq 0.05$) increased concentration (by about 11% each with soil and foliar application of N) and contents (by about 23 and 16% respectively with soil and foliar-applied N) of protein in grains (Table 3). The contents of protein in grains were significantly ($P \leq 0.05$) less with soil-applied N than its foliar application.

Grain zinc

Grain Zn concentration ranged from 19 to 30 mg kg⁻¹ among twelve treatments comprising of all combinations of four Zn and three N levels (Figure 1). Grain Zn concentration was significantly ($P \leq 0.05$) influenced by main and interactive effects of N and Zn applications at panicle initiation (Table 1). At each level of N, application of Zn by any method significantly ($P \leq 0.05$) increased grain Zn concentration over control Zn (Figure 1). Similarly, at each level of N, the maximum increase in grain Zn concentration was achieved with combined soil + foliar Zn (36 to 54%) followed by foliar Zn alone (27 to 45%) and then soil Zn alone (9 to 15%).

At control level of Zn, the application of N, either to soil or on foliage, had a non-significant effect on grain Zn concentration over control level of N (Figure 1). Soil N increased grain Zn concentration over control level of N only when combined with soil Zn application. Foliar N

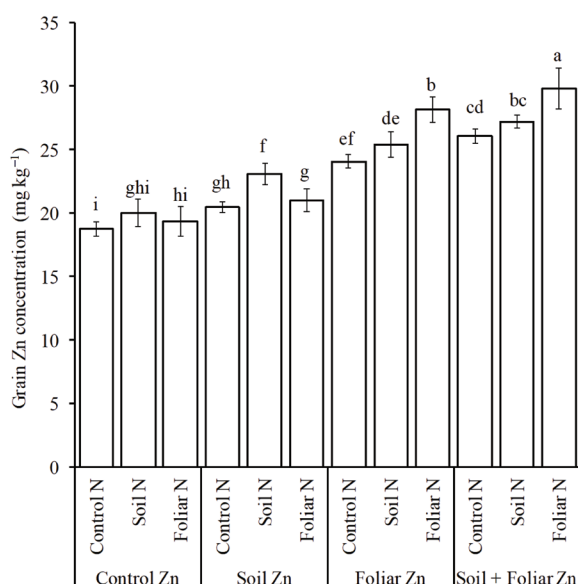


Figure 1. Concentration of zinc (Zn) in grains of rice grown in pots and fertilised, at panicle emergence stage, with all possible combinations of four Zn levels [control, soil application (6 mg Zn kg^{-1}), foliar spray ($2 \times 0.2\% \text{ w/v Zn}$) and soil + foliar application] and three nitrogen (N) levels [control, soil application (20 mg N kg^{-1}) and foliar spray ($0.5\% \text{ w/v N}$)]. Error bars are of standard deviations. Different letters on the bars indicate significant ($P \leq 0.05$) differences based on Tukey's HSD.

increased grain Zn concentration over control level of N only when combined with treatments having foliar Zn application. Maximum grain Zn concentration was achieved with foliar N application to pots supplied with combined soil + foliar Zn applications.

Only main effects of N and Zn were significant ($P \leq 0.05$) for grain Zn contents (Table 4). As compared to control level of Zn, application of Zn by either method increased grain Zn contents producing maximum value with soil + foliar-applied Zn. Similar to grain Zn concentration, Zn contents were mainly increased by the treatments having foliar Zn applications; an increase of 38 and 53% respectively was recorded by foliar alone and soil + foliar applications of Zn over control level of Zn.

DISCUSSION

Soil Zn application, and both soil and foliar N applications at panicle initiation significantly ($P \leq 0.05$) increased straw and grain yield of rice (Table 2). This is because plants require a continuous supply of most of the nutrients throughout their life cycle. The level of yield response by a crop, however, is dependent on the rate, source, method and time of fertilizer application (18). In the present study, Zn and N were applied at panicle initiation on 50% of main-tillers. During treatment application, therefore, many sub-tillers might have been in vegetative growth. A

growth response was therefore expected especially due to the reason that soil was low in plant-available Zn and previously applied N might have already been used by plant or lost in the environment. Moreover, N application at panicle initiation stage of rice increases grain yield by increasing grain weight in inferior spikelets (19). However, application of N at late vegetative stage delays maturity and increases chances of lodging (20). Therefore, field optimisation of rate and exact time of N application must be studied for economic yield returns along with quality grains.

Efforts have been done to increase protein and mineral densities in rice grains by conventional plant breeding and genetic engineering. Along with environmental protection and higher yields, agronomic fertiliser management is also meant to produce nutritious plant-based foods. For example, N fertilisation increased Zn and protein contents in rice grains along with an increase in grain yield (21). In the present study, soil N fertilisation at panicle initiation also played a positive role in increasing grain protein concentration and contents in rice (Table 3). Application of N to cereals also increases grain accumulation of Zn by an increased sink in the form of water-insoluble proteins (22, 23). Other reasons for increased grain Zn concentration by N application include increased uptake by the intensified root system, and better translocation and remobilisation of Zn towards grains (24, 25). Therefore, a suitable quantity of N fertilizer at the flowering stage of rice is an important measure to obtain higher grain protein and grain Zn concentration.

Similar to N, the application of Zn also increases protein and Zn concentration in rice grains. Soil application, seed priming and foliar application are recommended to both flooded and direct seeded rice for this purpose (9).

Table 4. Contents of zinc (Zn) in grains of rice grown in pots and fertilised, at panicle emergence stage, with all possible combinations of four Zn levels and three nitrogen (N) levels

Treatment levels	Grain zinc content (mg pot^{-1})
Main effect of Zn levels	
Control Zn	108±12 D
Soil Zn (6 mg Zn kg^{-1})	128±10 C
Foliar Zn ($0.2\% \text{ Zn w/v}$)	149±15 B
Soil Zn (6 mg Zn kg^{-1}) + Foliar Zn ($0.2\% \text{ Zn w/v}$)	165±19 A
Main effect of N levels	
Control N	123±19 B
Soil N ($20 \text{ mg kg}^{-1} \text{ soil}$)	146±22 A
Foliar N ($0.5\% \text{ N w/v}$)	142±31 A

Means ± standard deviations; Separately for each main effect, different letters indicate significant ($P \leq 0.05$) differences based on Tukey's HSD.

Zinc influences several physiological functions like synthesis, integrity and functioning of proteins in plants (26). Moreover, Zn application might influence the uptake and translocation of N in rice plants (13). Therefore, Zn application increased concentration and contents of protein in rice grains (Table 4).

From soil-applied Zn fertiliser, soil Zn^{2+} ion rapidly forms precipitates of zinc carbonate (ZnCO_3) under alkaline calcareous soils (27). Therefore, recovery of soil-applied Zn is limited in high pH calcareous soils. Moreover, the availability of applied Zn decreases over time (28). Therefore, late applications of Zn, especially in foliar form, are ideal for efficient uptake by plants. Late soil applications are also justifiable as a major fraction of Zn in grains is as a result of Zn uptake by roots after flowering stage (29). Soil Zn application combined two foliar Zn applications at flowering are suggested by researchers for effective Zn biofortification in rice (13, 30). Foliar Zn sprays are important as Zn is not fixed in soils and it has to travel little from leaf to grain as comparison to the soil where it travels from roots to grain (31).

In the present study, soil N increased grain Zn concentration if combined soil Zn and foliar N application increased grain Zn concentration if combined with treatments having foliar Zn (Figure 1). Zinc translocate through xylem as Zn-phytosiderophore or Zn-amino acid and in phloem as Zn-nicotinamine complexes (32). These organic compounds have N as their integral part. For N fertilized soils, therefore, the cotransport of Zn with N seems important for increased grain Zn accumulation by N application.

CONCLUSIONS

At panicle initiation, application of Zn and N increased concentrations of protein and Zn in rice grains. Results suggested that grain Zn concentration is significantly ($P \leq 0.05$) increased over respective control when both Zn and N were combined to soil or on foliage. This suggests that the role of N in Zn uptake, absorption, translation, and remobilisation is important for cotransport of Zn with N. Foliar + soil Zn when combined with foliar N increased grain Zn concentration to an acceptable level of Zn biofortification in rice.

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DECLARATION

Authors declare no competing interests.

REFERENCES

1. VANLAUWE B, SIX J, SANGINGA N, ADESINA AA 2015 Soil fertility decline at the base of rural poverty in sub-Saharan Africa. *Nat Plants* 1(7): 15101. <https://doi.org/10.1038/nplants.2015.101>
2. HUSSAIN S, MAQSOOD MA, RENGEL Z, KHAN MK 2012 Mineral bioavailability in grains of Pakistani bread wheat declines from old to current cultivars. *Euphytica* 186(1): 153–163. <https://doi.org/10.1007/s10681-011-0511-1>
3. FAN M-S, ZHAO F-J, FAIRWEATHER-TAIT SJ, POULTON PR, DUNHAM SJ, MCGRATH SP 2008 Evidence of decreasing mineral density in wheat grain over the last 160 years. *J Trace Elem Med Biol* 22(4): 315–24. <https://doi.org/10.1016/j.jtemb.2008.07.002>
4. BOUIS HE, SALTZMAN A 2017 Improving nutrition through biofortification: A review of evidence from HarvestPlus, 2003 through 2016. *Glob Food Sec* 12(January): 49–58. <https://doi.org/10.1016/j.gfs.2017.01.009>
5. DUFFNER A, HOFFLANDE E, STOMPH TJ, MELSE-BOONSTRA A, BINDRABAN PS 2014 Elimination zinc deficiency in rice-based systems (VFRC Report 2014/2), Washington, D.C.
6. ALLOWAY B 2009 Soil factors associated with zinc deficiency in crops and humans. *Environ Geochem Health* 31(5): 537–548. <https://doi.org/10.1007/s10653-009-9255-4>
7. QASWAR M, HUSSAIN S, RENGEL Z 2017 Zinc fertilisation increases grain zinc and reduces grain lead and cadmium concentrations more in zinc-biofortified than standard wheat cultivar. *Sci Total Environ* 605–606: 454–460. <https://doi.org/10.1016/j.scitotenv.2017.06.242>
8. RAM H, RASHID A, ZHANG W, DUARTE AP, PHATTARAKUL N, SIMUNJI S, KALAYCI M, FREITAS R, RERKASEM B, BAL RS, MAHMOOD K, SAVASLI E, LUNGU O, WANG ZH, BARROS VLNP DE, MALIK SS, ARISOY RZ, GUO JX, SOHU VS, ZOU CQ, CAKMAK I 2016 Biofortification of wheat, rice and common bean by applying foliar zinc fertilizer along with pesticides in seven countries. *Plant Soil* 403(1): 389–401. <https://doi.org/10.1007/s11104-016-2815-3>
9. FAROOQ M, ULLAH A, REHMAN A, NAWAZ A, NADEEM A, WAKEEL A, NADEEM F, SIDDIQUE KHM 2018 Application of zinc improves the productivity and biofortification of fine grain aromatic rice grown in dry seeded and puddled transplanted production systems. *F Crop Res* 216: 53–62. <https://doi.org/10.1016/j.fcr.2017.11.004>
10. IMRAN M, KANWAL S, HUSSAIN S, AZIZ T, MAQSOOD MAMA 2015 Efficacy of zinc application methods for concentration and estimated bioavailability of zinc in grains of rice grown on a calcareous soil. *Pakistan J Agric Sci* 52(1): 169–175
11. PERSSON DP, BANG TC DE, PEDAS PR, KUTMAN UB, CAKMAK I, ANDERSEN B, FINNIE C, SCHJOERRING JK, HUSTED S 2016 Molecular speciation and tissue compartmentation of zinc in durum wheat grains with contrasting nutritional status. *New Phytol* 211(4): 1255–1265. <https://doi.org/10.1111/nph.13989>
12. ERENOGLU EB, KUTMAN UB, CEYLAN Y, YILDIZ B, CAKMAK I 2011 Improved nitrogen nutrition enhances root uptake, root-to-shoot translocation and remobilization of zinc (^{65}Zn) in wheat. *New Phytol* 189(2): 438–448. <https://doi.org/10.1111/j.1469-8137.2010.03488.x>
13. GUO JX, FENG XM, HU XY, TIAN GL, LING N, WANG JH, SHEN QR, GUO SW 2016 Effects of soil zinc availability, nitrogen fertilizer rate and zinc fertilizer application method on zinc biofortification of rice. *J Agric Sci* 154(4): 584–597. <https://doi.org/10.1017/S0021859615000441>
14. SPARKS DL, PAGE AL, SUMNER ME, TABATABAI MA, HELMKE PA 1996 *Methods of Soil Analysis – Part 3. Chemical Methods*, Soil Science Society of America, Madison, USA

15. KLUTE A 1986 Methods of soil analysis. Part 1. Physical and mineralogical methods., American Society of Agronomy, Inc., Madison. <https://doi.org/10.2136/sssabookser5.1.2ed.c9>
16. JONES JB, CASE VW 1990 Sampling, handling and analyzing plant tissue samples, In: Westerman R L (ed) Soil Testing and Plant Analysis, Soil Science Society of America, Madison, USA, p 389
17. GINKEL JH VAN, SINNAEVE J 1980 Determination of total nitrogen in plant material with Nessler's reagent by continuous-flow analysis. *Analyst* 105(1257): 1199. <https://doi.org/10.1039/an9800501199>
18. XU X, HE P, QIU S, PAMPOLINO M F, ZHAO S, JOHNSTON AM, ZHOU W 2014 Estimating a new approach of fertilizer recommendation across small-holder farms in China. *F Crop Res* 163: 10–17. <https://doi.org/10.1016/j.fcr.2014.04.014>
19. JIANG Q, DU Y, TIAN X, WANG Q, XIONG R, XU G, YAN C, DING Y 2016 Effect of panicle nitrogen on grain filling characteristics of high-yielding rice cultivars. *Eur J Agron* 74: 185–192. <https://doi.org/10.1016/j.eja.2015.11.006>
20. CORBIN JL, ORLOWSKI JM, HARRELL DL, GOLDEN BR, FALCONER L, KRUTZ LJ, GORE J, COX MS, WALKER TW 2016 Nitrogen strategy and seeding rate affect rice lodging, yield, and economic returns in the midsouthern United States. *Agron J* 108(5): 1938–1943. <https://doi.org/10.2134/agronj2016.03.0128>
21. GU J, CHEN J, CHEN L, WANG Z, ZHANG H, YANG J 2015 Grain quality changes and responses to nitrogen fertilizer of japonica rice cultivars released in the Yangtze River Basin from the 1950s to 2000s. *Crop J* 3(4): 285–297. <https://doi.org/10.1016/j.cj.2015.03.007>
22. XUE Y-F F, EAGLING T, HE J, ZOU C-Q Q, MCGRATH SP, SHEWRY P R, ZHAO F-J J, MCGRATH SP, SHEWRY PR, ZHAO F-J J 2014 Effects of nitrogen on the distribution and chemical speciation of iron and zinc in pearling fractions of wheat grain. *J Agric Food Chem* 62(20): 4738–4746. <https://doi.org/10.1021/jf500273x>
23. KUTMAN UB, YILDIZ B, CAKMAK I 2011 Effect of nitrogen on uptake, remobilization and partitioning of zinc and iron throughout the development of durum wheat. *Plant Soil* 342(1–2): 149–164. <https://doi.org/10.1007/s11104-010-0679-5>
24. NIE Z, ZHAO P, WANG J, LI J, LIU H 2017 Absorption kinetics and subcellular fractionation of zinc in winter wheat in response to nitrogen supply. *Front Plant Sci* 8: 1435. <https://doi.org/10.3389/fpls.2017.01435>
25. ZHAO P, YANG F, SUI F, WANG Q, LIU H 2016 Effect of nitrogen fertilizers on zinc absorption and translocation in winter wheat. *J Plant Nutr* 39(9): 1311–1318. <https://doi.org/10.1080/01904167.2015.1106560>
26. BROADLEY M, WHITE P, HAMMOND J 2007 Zinc in plants. *New Phytol* 677–702. <https://doi.org/10.1111/j.1469-8137.2007.01996.x>
27. KAMALI S, RONAGHI A, KARIMI N 2010 Zinc Transformation in a Calcareous Soil as Affected by Applied Zinc Sulfate, Vermicompost, and Incubation Time. *Commun Soil Sci Plant Anal* 41(19): 2318–2329. <https://doi.org/10.1080/00103624.2010.508096>
28. HUSSAIN S, MAQSOOD MA, RAHMATULLAH 2011 Zinc release characteristics from calcareous soils using diethylenetriaminepentaacetic acid and other organic acids. *Commun Soil Sci Plant Anal* 42(15): 1870–1881. <https://doi.org/10.1080/00103624.2011.587571>
29. JOHNSON-BEEBOUT SE, GOLORAN JB, RUBIANES FHC, JACOB JDC, CASTILLO OB 2016 Zn uptake behavior of rice genotypes and its implication on grain Zn biofortification. *Sci Rep* 6: 38301. <https://doi.org/10.1038/srep38301>
30. SAHA S, CHAKRABORTY M, PADHAN D, SAHA B, MURMU S, BATABYAL K, SETH A, HAZRA GC, MANDAL B, BELL RW 2017 Agronomic biofortification of zinc in rice: Influence of cultivars and zinc application methods on grain yield and zinc bioavailability. *F Crop Res* 210: 52–60. <https://doi.org/10.1016/j.fcr.2017.05.023>
31. WISSUWA M, ISMAIL AM, GRAHAM RD 2008 Rice grain zinc concentrations as affected by genotype, native soil-zinc availability, and zinc fertilization. *Plant Soil* 306(1–2): 37–48. <https://doi.org/10.1007/s11104-007-9368-4>
32. OLSEN L I, PALMGREN MG 2014 Many rivers to cross: the journey of zinc from soil to seed. *Front Plant Sci* 5(February): 30. <https://doi.org/10.3389/fpls.2014.00030>